IJP 02919

Ketobemidone prodrugs for buccal delivery: Prediction of the extent of saliva-catalyzed hydrolysis of various ester prodrugs under simulated in vivo conditions

Laila Bach Hansen^a, Lona Louring Christrup^a and Hans Bundgaard^b

Departments of a Pharmaceutics and b Pharmaceuttcal Chemtstry, The Royal Damsh School of Pharmacy, Copenhagen (Denmark)

(Recewed 6 April 1992) (Accepted 8 May 1992)

Key words: Ketobemidone; Prodrug; Saliva enzyme-catalyzed hydrolysis; Ester; Buccal delivery

Summary

As part of studies aiming at developing a ketobemidone prodrug suitable for buccal or sublingual administration, the potential ~mpact of salwa enzyme-catalyzed hydrolysis of various ester prodrugs was assessed. The hydrolysis of three ketobemidone esters in human whole saliva, obtained under conditions ensuring maximal esterase activity, was studied as a function of ester concentration at 37°C The kinetics of hydrolysis could be accounted for in terms of the Michaelis-Menten equation and the rate parameters K_m and V_{max} were determined. Due to the occurrence of zero-order kinetics at pharmacologically relevant prodrug concentrations, degradation of the esters by saliva enzymes was predicted to occur to only a minor extent $(1-6%)$ under conditions similar to those prevailing in vivo after administration of buccal or sublingual tablets of the esters. The mode of administration of tablets for use in the mouth and their rate of disintegration were shown to have some influence on the rate of salwa secretion and hence on sahva esterase activity but not to an extent compromising the efficient buccal or subhngual delivery of the ketobemldone prodrugs.

Introduction

Ketobemidone, 1-[4-(3-hydroxyphenyl)- 1-methyl-4-piperidyl]-l-propanone (I), is a strong narcotic analgesic equipotent with morphine (Eddy et al., 1957; Anderson et al., 1986) which has been used clinically for more than 40 years, especially in Scandinavia and in some other European countries. The drug is usually administered perorally or rectally but its bioavailability is incomplete and variable due to pronounced first-pass metabolism (Bondesson et al., 1980; Anderson et al., 1981, 1986).

Studies have been initiated in our laboratories to develop a ketobemidone preparation suitable for buccal or sublingual administration in order to improve its bioavailability (through avoidance of the first-pass effect) and achieve a more rapid onset of action relative to the peroral or rectal preparations. Initial studies showed that ketobemidone did not penetrate the oral mucosa, presumably due to its poor lipophilicity, and therefore, we have prepared a number of carboxylate

Correspondence to. L.L. Christrup, The Royal Danish School of Pharmacy, Department of Pharmaceutics, 2 Universitetsparken, DK-2100 Copenhagen, Denmark.

or carbonate ester prodrugs at the phenolic hydroxyl group in the drug (Hansen et al., 1991). These derivatives were found to be more lipophilic than the parent drug and to be rapidly and quantitatively hydrolyzed to ketobemidone in the presence of human plasma. However, most of the esters were also rapidly hydrolyzed when incubated in whole human saliva at 37°C (Hansen et al., 1991). Human saliva contains a variety of esterases (Chauncey et al., 1954, 1957; Burstone, 1956; Chauncey, 1961; Lindqvist and Augustinsson, 1975; Tan, 1976; Lindqvist et al., 1977) and the last finding thus makes it important to consider the possible impact of saliva enzyme-catalyzed degradation of the ester prodrugs in the development of buccal or sublingual preparations.

In the accompanying paper (Hansen et al., 1992), various factors influencing the esterase activity of human saliva have been studied. A most important factor was shown to be the salivation flow rate in that the esterase activity increased greatly with increasing salivary flow up to a rate of about 0.9 ml min⁻¹. Different stimuli such as paraffin chewing and sucking calcium phosphate tablets, acid drops or sour lemon candy result in increased salivary flow (Kerr, 1961; Zickert, 1974; Minaguchi et al., 1988). It may therefore be imagined that the type of device used for a buccal or sublingual preparation can effect the salivary flow and hence the salivary esterase activity.

The primary objective of the present study was to provide information on the stability of various ketobemidone ester prodrugs $(II-IV,$ Scheme 1) in human saliva at various prodrug concentrations including clinically relevant concentrations. In addition, the effect of slowly and rapidly disintegrating buccal or sublingual tablets with and without taste on the salivary flow was examined.

Materials and Methods

Chemicals

Ketobemidone hydrochloride was obtained from H. Lundbeck A/S, Copenhagen, Denmark. The ketobemidone esters II-IV were prepared as previously described (Hansen et al., 1991). The

ester II was used as the fumarate salt whereas the other two esters were in the form of hydrochloric acid salts.

Influence of buccal / sublingual tablet composttion and application site on salivary flow

Buccal/sublingual tablets $(120-140 \text{ mg})$ having either fast or slow disintegration rates were used in the study. Each type of tablets was made with or without taste (0.5 mg of) per tablet). The rapidly disintegrating sublingual tablets were placed under the tongue whereas the slowly disintegrating tablets were placed either sublingually or buccally (between the upper lip and gum). Control experiments without tablet administration were also performed. Two female subjects (age 29 and 36 years) participated in the

study. After application of the buccal/sublingual tablet, whole saliva was collected by expectoration into centrifuge tubes for every 2 min over a period of 20 min; whereupon any residues of the tablet were removed. Saliva collection was continued for 10 min. Seven experiments were performed per day with each subject, interrupted by 30 min intervals in which water but not food was allowed. The experiments were carried out with three replications.

Collection of saliva for stability experiments

Whole saliva, stimulated by normal frequent paraffin chewing giving a salivary flow of 1.0-1.2 ml min^{-1}, was collected from subject 1 before breakfast and following a fasting period of 10 h. When chewing the paraffin was moved from one side of the mouth to the other in order to activate all salivary glands (Kerr, 1961). Brushing of teeth was only allowed with toothpaste without fluoride and performed at least 1 h prior to saliva collection, since it has been found that the esterases in human whole saliva can be inhibited by several of the commonly used dental materials (Lindquist et al., 1980).

Hydrolysis studies in human sahca

The hydrolysis of the ketobemidone esters II-IV was studied in fresh whole human saliva obtained as described above. The initial concentrations of the esters were in the range 1.8×10^{-4} - 1.5×10^{-2} M and the initial rate method was used to determine the kinetics of hydrolysis. The reactions were initiated by adding $25-500 \mu$ l of a stock solution of the esters in water or wateracetonitrile to 5.00 ml of whole human saliva. The solutions were kept in a water-bath at $37.0 \pm$ 0.2°C, and at appropriate intervals, 250 μ l samples were withdrawn and added to 500 μ l of a 2% solution of zinc sulphate in acetonitrile-water $(1:1 \text{ v/v})$ in order to deproteinize the saliva. After mixing and centrifugation for 3 min at 13000 rpm, 20 μ l of the clear supernatant was analyzed for ketobemidone by a reversed-phase HPLC procedure described previously (Hansen et al., 1991). The detection limit for ketobemidone was 0.1 μ g ml⁻¹, corresponding to 4×10^{-7} M. The initial rate of ester hydrolysis was determined from the slopes of linear plots of amount of ketobemidone formed against time. In all experiments, less than 5% of the ester was hydrolyzed, implying an almost constant substrate concentration during the runs.

Results and Discussion

Variation in salivary flow wtth composition of buccal / sublingual tablets

The influence of slowly and rapidly disintegrating tablets with or without 0.5 mg of peppermint oil as flavouring agent on the salivary flow was tested in two subjects after buccal or sublingual administration. The tablets with a fast disintegration rate were only administered sublingually. The results are shown in Fig. 1.

Significant differences ($p < 0.001$) in the salivary flow were found between the two subjects at all sampling times. A marked interindividual variation in paraffin chewing-stimulated salivation rates has previously been observed (Bertram, 1967; Heintze et al., 1983; Barenthin and Johnson, 1986). No significant effect of peppermint oil was observed and therefore, the data in Fig. 1 for each application site include both tablets with and without a content of this flavouring agent. Significant differences were found between the application sites at 2-28 min for subject 1 and at 2-6 min for subject 2 ($p < 0.05$), the highest salivary flow occurring after administration of rapidly disintegrating sublingual tablets. The disintegration time of these tablets in vivo was 1.5- 3.5 min.

The weight loss of the buccal/sublingual tablets determined 20 min after their administration is listed in Table 1. Despite equal salivary flow rates after administration of slowly disintegrating buccal and sublingual tablets, the buccal tablets showed a smaller loss. Administration af slowly disintegrating or dissolving buccal tablets may be preferred over that of slowly disintegrating or dissolving sublingual tablets because of the prolonged retention time in the oral cavity and also because the presence of a buccal tablet is not noticeable to the patients after a short time (Livingstone and Livingstone, 1989).

Stabihty of ketobemidone prodrugs tn human saliva

The ketobemidone esters II-IV have previously been found to undergo an enzyme-catalyzed hydrolysis to yield ketobemidone in quantitative amounts when incubated in whole human saliva at 37°C (Hansen et al., 1991). At an initial concentration of 10^{-4} M the hydrolysis proceeded according to first-order kinetics with half-lives of 295 min (II), 5.1 min (III) and 4.9 min (IV), respectively. The corresponding half-lives for the hydrolysis in 80% human plasma were 1.8 min (II) , 0.03 min (III) and 0.04 min (IV) whereas those in buffer solutions of pH 7.4 and at 37°C exceeded 50-100 h (Hansen at al., 1991).

At higher initial ester concentrations the rate of hydrolysis of the esters in saliva changed to follow zero-order kinetics which is typical for enzyme-catalyzed reactions following Michaelis-Menten kinetics in which the initial substrate concentration is higher than the Michaelis constant K_m (Segel, 1975). The initial rates of formation of ketobemidone (V_0) from the esters were determined at different initial ester concentrations (Figs 2 and 3) and shown to conform to the Michaelis-Menten equation:

$$
V_0 = \frac{V_{\text{max}} S_0}{K_{\text{m}} + S_0}
$$
 (1)

where V_{max} is the maximum rate of substrate consumption, $K_{\rm m}$ represents the Michaelis constant and S_0 is the initial (and almost constant) substrate concentration. The Michaelis-Menten parameters K_m and V_{max} were determined from Lineweaver-Burk plots of Eqn 1 (an example is shown in Fig. 4) and are listed in Table 2.

It should be noted that the saliva used in the kinetic runs was obtained under conditions favouring a high esterase activity (Hansen et al., 1992).

Inspection of the data in Table 2 shows that the ester III is the best substrate, having the greatest V_{max} value and the lowest K_{m} value (tightest binding). The most stable compound is the 3,3-dimethylbutyrate ester (II) which may be ascribed to steric hindrance by the bulky alkyl side chain. The low reactivity of ester II may also partly be ascribed to a decrease in pH of the saliva upon addition of the ester which was used in the form of its fumarate salt. Thus, whereas addition of the esters III and IV (hydrochloric acid salts) at a concentration of 1.5×10^{-2} M to saliva did not change the pH from that in a saliva sample without ester (pH 6.9–7.0), addition of ester II at a similar concentration afforded a pH value of 4.7. We have previously found that the esterase activity of saliva at pH 4.5 is about 1.6-times lower than at pH 7 (Hansen et al., 1992).

The rate data obtained along with Eqn 1 permit the calculation of the extent of saliva-catalyzed degradation of the ketobemidone esters under clinically relevant conditions. Thus, the

TABLE 1 *Weight loss of sublingual or buccal tablets withdrawn 20 min after administration to two subjects (mean* \pm *SD, n = 3)*

Type of tablet (disintegration)	Flavouring agent added ^a	Site of administration	Weight loss $(\%)$		
			Subject 1	Subject 2	
Fast ^b	yes	sublingual	100	100	
Fast ^b	no	sublingual	100	100	
Slow	ves	sublingual	$35 + 11$	59 ± 14	
Slow	no	sublingual	44 ± 7	$58 + 16$	
Slow	yes	buccal	$9 + 1$	16 ± 3	
Slow	no	buccal	11 ± 3	13 ± 3	

^a 0.5 mg peppermint oil/tablet.

 b Time of disintegration was 1.5-3.5 min</sup>

Fig 1. Plots of cumulative saliva secretion against time after administration of rapidly (Δ) and slowly (\square) disintegrating sublingual tablets, and administration of slowly disintegrating buccal tablets (\circ) to two subjects; (\bullet) control data (i.e., saliva secretion when no tablets were gwen) Error bars represent S.D $(n = 3-6)$.

amount of ester degraded in 5 min can be expressed as:

$$
\text{Degraded ester } (\%) = \frac{V_{\text{max}} \times 5}{K_{\text{m}} + S_0} \times 100 \tag{2}
$$

The results of such calculations as a function of saliva volume are shown in Fig. 5 for ester pro-

Fig. 2. Plots showing the rate of formation of ketobemldone upon hydrolysis of compound IV m whole human sahva (37°C) at initial concentrations of compound IV of 4.0×10^{-4} M (\bullet), 1.3×10^{-3} M (\triangle) and 13×10^{-2} M (\blacksquare)

Fig 3. The effect of substrate concentration on the initial rate of the sahva catalyzed hydrolysis of compound $II(\bullet)$, III (\blacksquare) and IV (\triangle). The full lines have been calculated from Eqn 1 and the K_m and V_{max} values in Table 2.

TABLE 2

Michaelis-Menten parameters K_m *and* V_{max} *for the hydrolysis of varmus ketobemMone ester prodrugs at 37°C m human whole sabra*

Compound	ռա (M)	$V_{\rm max}$ $(M \text{ min}^{-1})$
п	29×10^{-3}	12×10^{-6}
ш	5.5×10^{-4}	23.9×10^{-6}
ΓV	7.4×10^{-4}	9.8×10^{-6}

Fig 4 Lmeweaver-Burk plot of the rate data for the sahva catalyzed hydrolysis of ketobemldone ester IV

drug doses corresponding to 5 or 10 mg ketobemidone hydrochloride (normal doses). In the calculations the assumption was made that the doses

Fig 5. Plots of % ketobemldone prodrug hydrolyzed during incubation for 5 min in human saliva against saliva volume at prodrug concentrations equivalent to 5 mg $($ ----) or 10 mg \cdot) ketobemidone hydrochloride. Ester II (\bullet), III (\blacksquare) and $\mathbf{IV}(\blacktriangle)$ The curves were calculated from Eqn 2

apphed were dissolved immediately and homogenously distributed in the saliva, the volume of which remained constant during the 5 min period. The amount of saliva normally secreted during 5 min is in the range of 1.5-4 ml. As can be seen from Fig. 5, hydrolysis of a ketobemidone ester prodrug by saliva enzymes at pharmacologically relevant concentrations is only expected to occur to a very minor extent which in turn is due to the occurrence of zero-order kinetics at these ester concentrations.

Acknowledgements

This study was supported by grants from H. Lundbeck A/S, Copenhagen, Denmark and the Danish Research Academy, Arhus, Denmark.

References

- Anderson, P, Arnér. S., Bondesson, U., Boréus, LO and Hartvig, P., Chnical pharmacokinetics of ketobemidone Its bloavallability after rectal administration *Eur J Chn Pharmacol,* 19 (1981) 217-223.
- Anderson, P, Arnér, S, Bondesson, U, Boréus, LO and Hartvig, P, Pharmacokinetics of ketobemidone. In Foley, K.M. and Inturrisi, C.E. (Eds), *Advances in Pain Research and Therapy,* Vol. 8, Raven, New York, 1986, pp. 171-178.
- Barenthin, I and Johnson, G., Frequency of low values of stimulated sahva flow rate, Dentobuff and Dentocult m routine use *Swed Dent. J,* 10 (1986) 207-212.
- Bertram, U., Xerostomia, clinical aspects, pathology and pathogenesis. *Acta Odontol Scand*, 25 (Suppl 49) (1967) 12-30
- Bondesson, U, Arnér, S, Anderson, P, Boréus, L.O and Hartvig, P, Clinical pharmacokinetics and oral bioavailability of ketobemldone *Eur J. Chn. Pharmacol,* 17 (1980) 45-50
- Burstone, M.S., Esterase of the salivary glands *J Histochem Cytochern,* 4 (1956) 130-139.
- Chauncey, H H., Lionetti, F and Lisanti, V.F, Enzymes of human salıva II Parotid saliva total esterases *J Dent Res,* 36 (1957) 713-716
- Chauncey, H.H, Lionetti, F, Winer, R A. and Lisanti, V F Enzymes of human saliva. I The determination, distribution, and ongm of whole salwa enzymes. *J Dent Res,* 33 (1954) 321-334
- Chauncey, H.H, Salwary enzymes *J Am Dent Assoc,* 63 (1961) 360-368
- Eddy, N.B, Halbach, H and Braenden, O.J,, Synthetic substances with morphine-hke effect Climcal experience po-

tency, side-effects, addiction liability *Bull* WHO, 17 (1957) 569-863.

- Hansen, L.B., Christrup, L.L and Bundgaard, H, Ketobemidone prodrugs for buccal dehvery *Acta Pharm Nord, 3* (1991) 77-82.
- Hansen, L B, Christrup, L.L and Bundgaard, H., Salivacatalyzed hydrolysis of a ketobemidone ester prodrug-Factors influencing human salivary esterase activity. *Int J Pharm,* 88 (1992) 221-227
- Heintze, U., Birkhed, D and Bjorg, H., Secretion rate and buffer effects of resting and stimulated whole sahva as a function of age and sex. *Swed. Dent* J, 7 (1983) 227-238.
- Kerr, *A C The Physiological Regulation of Sahcary Secretions m Man A Study of the Response of Human Salwary Glands to Reflex Stimulation,* Pergamon, Oxford, 1961, Ch 6, pp 48-60
- Lmdqvist L. and Augustinsson, K.-B, Esterases in human sahva *Enzyme,* 20 (1975) 277-291.
- Lindqvist, L., Bartfai, T, Berg J.-O and Blomlof, L., In vivo and m vitro studies of inhibitory effects of restorative dental materials on sahvary esterases. Scand. *J Dent Res •* 88, I (1980) 229-235
- Lmdqvlst, L, Nord, C.-E. and Soder, P.-O, Origin of esterases in human whole saliva *Enzyme,* 22 (1977) 166-175
- Livingstone, C and Livingstone, D., Drug dehvery (4) Buccal and rectal routes *Pharm J,* 242 (1989) 68-69
- Minaguchi, K, Madapallimattan, G and Bennick, A, The presence and origin of phosphopeptides in human saliva *Btochem J,* 250 (1988) 171-177
- Segel, I.H., *Enzyme Kinetics*, Wiley, New York, 1975, pp. 18-57.
- Tan, S.G, Human saliva esterases' Genetic studies *Hum Hered,* 26 (1976) 207-216
- Zickert, I, Sahvsekretlon vld ohka Iokala stimuli *TandldkartMnmgen,* 24 (1974) 1439-1441.